# Impact of Cassava Mill Effluent (CME) on Soil Physicochemical and Microbial Community Structure and Functions

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#### Abstract

Impact of Cassava Mill Effluent (CME) on soil physicochemical parameters microbial diversity and enzyme activities were analyzed in Akacze, an agrarian community of Ebonyi State Nigeria. Results showed change in temperature (28.6-326°C), pH (7.2-10.3) and TOC (24.2-41.3mg/g). Highest values were obtained nearest the waste pit while control soil had the least values. Total P, total N and C/N ratio were ilso lowest nearest the pit but cyanogenic potential was highest nearest pit. The soil heavy and trace metals Hg. Cr, Pb andCd) were not affected except Ca which was highest near the pit and decreased away from it. Total heterotrophic bacteria had a count range of 3.7x10<sup>4</sup>-6.6x10<sup>6</sup> cfu/g, lipolytic bacteria, had 0.9x10<sup>1</sup>-24x10<sup>1</sup> cfu/g while nitrifying bacteria had 0.4-2.9x3.7x103 cfu/g. Phosphate-solubilizing bacteria had a range of 2.2x10<sup>2</sup>-2.4x10<sup>3</sup> cfu/g. In all cases, highest values were obtained 100m from the waste pit, followed by the control while the least was in the waste pit edge. Dehydrogenase activities followed the pattern of THB-highest in 100m, control, 10m, 5m and pit edge was the least. Urease had the opposite trend with highest nearest the pit (mg g-<sup>1</sup>24-<sup>1</sup>) and least in control (2.2). Acid phosphatase had decreasing activities towards the pit while alkaline phosphates had increasing activities in the same direction. This trend was pH dependent. Results indicated adverse effects of the CME on soil parameters and calls for regulations on the disposal of CME to avoid environmental degradation.

Kerwords: Cassava, effluent, Bacteria, Enzyme, Soil, activities
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#### INTRODUCTION

Cassava (Manihot esculenta Crantz, synonymous with Manihot utilissima Rhol) belongs to the family Euphorbiaceae. It is mainly a food crop whose tubers are harvested between 7-13 months based on the cultivars planted (Cook, 1985; Tave, 1994). The tubers are quite rich in carbohydrates (85-90%) with very small amount of protein (1.3%) in addition to cyanogenic gloucoside (Linamarin and Lotaustiallin) (Nwabueze and Odunsi. 2007, Oyewole and Afolami, 2001). This high carbohydrate content makes cassava a major food item especially for the low income earners in most tropical countries especially Africa and Asia (Desse and Taye, 2001. Aderiye and Laleye, 2003).

The edible tubers are processed into various forms which include chips, pellets, cakes and flour. The flour could be fried to produce garri or steeped in water to ferment to produce foofoo when cooked (Oyewole

and Odunfa 1992). The production and consequent consumption of cassava have increased extensively in recent times. This increased utilization of processed cassava products has equally increased environmental pollution associated with the of the effluents. disposal The highly offensive odour emanating from fermenting effluent calls for regulation in the discharge of the waste generated (Akani, et al., 2006, Adewoye. et al. 2005).

In most areas, cassava mills are mainly on small scale basis, owned and managed by individuals who have no basic knowledge of environmental protection. Though on small scale basis, there are many of them, which when put together, create enormous impact on the environment. This work therefore on the environment. This work therefore was aimed at assessing the impact of cassava mill effluents on soil physicochemical and biological indices of

soil quality. This study will ascertain the health status of the cassava mill effluent impacted soil for human uses.

#### Materials and methods

The study area was Akaeze, a rural agrarian community of Ebonyi State, Nigeria. The community lies between latitude 5°53¹N and longitude 7°37¹. It is a tropical climate area with typical Guinea savanna vegetation characterized by very tall grasses and scattered trees. The people are mainly farmers with rice, cassava and yam being the main crops grown.

# Sample collection

Soil samples were collected using Shiprek soil augar disinfected with cotton wool soaked in 70% ethanol at 0-15 cm depth. Four sampling areas of the mill were chosen. The areas sampled were the pit edge, 5m, 10m and 100m away while the sample from 25om away served as the control. At each sampling distance, three samples were collected and pooled to give a composite sample for that particular area. Sterile universal bottles were used to collect the soil samples for microbiological and enzymatic studies. Clear amber coloured glass bottles, rinsed thrice in distilled water solution, were used for the soil samples for physicochemical properties analysis. The biological indices (enzymes and bacterial diversity) were analyzed within 2-4 hours of collection while the physicochemical parameters were determined within 12-24 hours of collection.

### Physicochemical properties

The soil temperature was determined *in-situ* using mercury in bulb thermometer inserted into the soil and allowed to stay for 5-10 minutes before reading was taken. The pH was determined using Jenway multipurpose tester (HANNA 1910 Modei) The organic matter content was estimated using the loss of ignition method (UNEP, 2004).

The heavy metals whose concentrations were analyzed include Pb, Cr, Cu and Cd while the trace elements were Mg, Ca, K and Na. The concentrations of these metals were determined using the AAS

method as described in APHA (1998) methods.

## Soil enzymes activities

The enzymes whose activities were assessed in this study include dehydrogenase, urease, and the phosphatase (acid and alkaline). The soil was dried at room temperature for 24 hours and passed through 0.5mm sieve. The sieved soil was used for the analysis.

Dehydrogenenase activity was determined by the method involving the reduction of TTC (Triphenyl tetrazolum chloride) to triphenyl formazon (TPF) after incubation of the TTC amended soil at 30°C for six hours as described by Alef (1995). activity was urease determined calorimetrically according to the method of Kandeler and Gerber (1988). The soil was incubated for 24 hours at 37°C with urea amendment and the result was expressed as NH3-N dry soil. Activities of both acid and alkaline phosphatases were determined as described by Tabatabai (1997) which involved the use of P-nitrophenyl phosphate and read at 410nm. While acid phosphatase activity was determined at pH 6.8, that of alkaline phosphatase was determined at Ph 11.5.

### Microbial diversity

The biological loads of various groups of bacterial species were determined using the culture techniques involving different selective culture media. Five different soil samples were analyzed for the bacteria diversity as described earlier in sample collection. Each sample was inoculated particular the media using the spread plate techniques as described by Chessbrough (2001) after ten-fold serial dilution. The bacterial groups were total bacteria heterotrophic (THB) presence was assessed with Tryptone Soy Agar, lipoytic bacteria with Tributyrin Agar and nitrifying bacteria (NB), assessed with modified mineral salt agar. The phosphatesolubilizing Agar was used to assess the phosphate solubilizing bacteria (PSB) as described in US patent (2003).

Conogenic potentials of the impacted

Cyanogenic poventials of impacted water was determined using modified pricate paper kits method as escribed by Bradburg et al., (1999). 2mls the water sample was put into 250ml emical flask containing 25ml distilled water. A strip of the test paper was soaked in the alkaline sodium picrate solution and find into the conical flask with the cork (stopper). The flask was allowed to stand at temperature for 18-24 hours. The was removed and eluted in 60ml The absorbance of the water was read # 540nm using spectrophotometer (Unican Hellosy England). The results recorded as mg HCN/L.

Statistical analysis

The results obtained in this study sere subjected to standard statistical malysis by the use of correlation analysis, sendard deviation and ANOVA. This was a determine the significance of the results.

Results The results of the physicochemical recerties are shown in Table 1. There were trends-either decreasing increasing gradients with distance away from the waste pit. The pH with a range of 12-10.3 temperature (28.6-32.6°C) and TOC (24.2-41.3 mg/g) were highest at the edge but lowest in the control soil sample. Total P (1.78-5.61mg/g) and total N (210-3.07 mg/g) had a similar trend. The 100m away and control soil samples values sere not statistically different (P =0.05) shere distance affected the other results significantly p=0.05). the C/N decreased significantly (p=0.05) away from edge of the waste pit (Table 1). The canogenic potential which was highest at the pit edge, decreased with distance away from the pit.

The results of the metal concentrations of the various heavy metals and trace metals are shown in Table 2.

There were no significant changes in the concentrations of the metals determined except in Ca (p=0.05). The highest calcium concentration was observed in the pit edge (2.1 mg/g) followed by the 5m (1.97mg/g) while the least was at the control soil (250m away).

In analysis of bacterial diversity, results obtained indicated that THB had the highest counts in all the soil samples analyzed. This group (THB) like all other bacterial groups, increased from the pit edge soil  $(3.7\times10^4 \text{ cfu/g lowest})$  to  $7.4\times10^6$ (highest) cfu/g in the 100m away soil. The 100m away soil value of THB was more than the control soil values. The LB  $(0.9x10^{1}-2.4x10^{3} \text{ cfu/g}) \text{ NB } (0.4x10^{1})$  $2.9 \times 10^3$  cfu/g) and PSB  $(2.2x10^2)$ 2.4x10<sup>4</sup>cfu/g) (Table 3) had prevalence with distance away from the pit. Though most of the bacterial groups had higher prevalence in the 100m soil than control soil sample, only that of THB was statistically significant (p=0.05).

Table 4 shows the soil enzymes activities of the five soil samples analyzed. Though all the enzymes showed some form of gradient in their activities, highest values were obtained at the 100m away soil samples. Urease (5.1 mg g<sup>-1</sup> 24<sup>-1</sup>) and alkaline phosphatase (6.7)umol-pnitrophenol) had their highest activities in gradually waste pit edge which decreased till the lowest in the control soil with 2.2 mg g-1 24h<sup>-1</sup> and 2.7 umol-pnitrophenol respectively. Dehydrogenase and acid phosphatase activities were highest in the 100m soil samples which gradually decreased with distance to the waste pit. The lowest values of dehydrogenase was 16.10 while the highest was 39.40. Lowest acid phosphatase value was 0.41 umolnitrop[henol, While dehydrogenase and acid phosphatase activities correlated positively with THB urease correlated negatively with THB but followed the trend of pH changes and total organic carbon

TABLE 1: Physicochemical Parameters of Soil

| Parameter          | Edge  | 5m    | 10m   | 100m | Control |
|--------------------|-------|-------|-------|------|---------|
| pH                 | 10.3  | 10.1  | 8.4   | 7.6  | 7.2     |
| Temp. °C           | 32.6  | 30.1  | 29.2  | 28.7 | 28.6    |
| TOC                | 41.3  | 35.0  | 30.8  | 24.4 | 24.2    |
| P                  | 1.78  | 2.12  | 2.73  | 5.19 | 5.61    |
| N                  | 2.10  | 2.16  | 2.28  | 3.91 | 3.07    |
| C/N Ratio          | 19.48 | 15.30 | 13.12 | 8.01 | 7.87    |
| Cyanogenic         | 5.21  | 4.66  | 3.22  | 0.92 | 0.62    |
| NH <sub>3</sub> -N | 5.2   | 4.3   | 2.7   | 0.79 | 0.72    |

Values are means of three times sampling

TABLE 2: Metal Concentration found in the Soil

| Edge  | 5m   | 10m  | 100m  | Control   |
|-------|--|--|---|---|
| 0.108 | 0.102  | 0.100  | 0.097   | 0.097   |
| 0.03  | 0.02   | 0.02   | 0.01  | 0.01  |
| 0.012 | 0.012  | 0.012  | 0.012   | 0.012   |
| 0.112 | 0.112  | 0.113  | 0.112   | 0.12  |
| 0.23  | 0.21   | .27  | 0.39  | 0.42  |
| 0.11  | 0.10   | 0.09   | 0.09  | 0.09  |
| 2.1   | 1.97   | 1.61   | 1.42  | 1.47  |
| 1.41  | 1.61   | 1.51   | 1.35  | 1.37  |
|       | 0.108<br>0.03<br>0.012<br>0.112<br>0.23<br>0.11<br>2.1 | 0.108       0.102         0.03       0.02         0.012       0.012         0.112       0.112         0.23       0.21         0.11       0.10         2.1       1.97 | 0.108       0.102       0.100         0.03       0.02       0.02         0.012       0.012       0.012         0.112       0.112       0.113         0.23       0.21       .27         0.11       0.10       0.09         2.1       1.97       1.61 | 0.108       0.102       0.100       0.097         0.03       0.02       0.02       0.01         0.012       0.012       0.012       0.012         0.112       0.113       0.112       0.112         0.23       0.21       .27       0.39         0.11       0.10       0.09       0.09         2.1       1.97       1.61       1.42 |

Values are means of three times sampling results

TABLE 3: Bacteria diversity (cfu/g) of the various soil Samples.

|       |                     | Locations           |                     |                     |                   |
|-------|---------------------|---------------------|---------------------|---------------------|-------------------|
| Group | Edge                | 5m                  | 10m                 | 100m                | Control           |
| THB   | $3.7 \times 10^4$   | $3.9 \times 10^4$   | 3.9x105             | $3.4 \times 10^6$   | $6.7 \times 10^6$ |
| LB    | $0.9 \times 10^{1}$ | $0.9 \times 10^{1}$ | $1.7 \times 10^{2}$ | $2.4 \times 10^3$   | $2.2 \times 10^3$ |
| NB    | $0.4 \times 10^{1}$ | $0.4 \times 10^{1}$ | $0.4 \times 10^{2}$ | $2.7 \times 10^{3}$ | $2.9 \times 10^3$ |
| PSB   | $2.2 \times 10^{2}$ | $2.4 \times 10^{2}$ | $2.1 \times 10^3$   | 2.x104              | $2.3 \times 10^4$ |

Values are means of three times sampling results

THB = Total Heterotrophic Bacteria

LB = Lipotytic Bacteria NB = Nitrifying Bacteria

PSB = Phosphate Solubilizing Bacteria

TABLE 4: Enzymatic activities evaluated in the soil samples

| Enzyme   | Edge    | 5m    | 10m   | 100m  | Control |
|--|---------|-------|-------|-------|---------|
| Dehydrogenase<br>mg g <sup>-1</sup> 6h <sup>-1</sup> | 16.10   | 20.30 | 28.75 | 39.40 | 37.50   |
| Urease mg g <sup>-1</sup> 24 <sup>1</sup>            | 5.1     | 4.8   | 3.6   | 2.4   | 2.2     |
| Acid phosphatase umol-<br>nitrophenol                | p- 0.41 | 1.2   | 2.7   | 3.3   | 3.6     |
| Alkaline phosphatase umol-<br>nitrophenol            | p- 6.7  | 6.4   | 5.1   | 3.1   | 3.4     |

# Discussion and conclusion

Results obtained in this study suggest adverse effects of the impact of cassava mill effluent on soil physicochemical and biological parameters. Analysis of these results showed that pH and temperature were hgighest very close to the waste pit. The same pit edge soil had highest organic carbon and cyanogenic potential but was lowest in P and Nwaugo et al. (2008a) and Akani et al. (2007) had reported similar situations in their studies. The TOC and low C/N ratio observed with increasing effects of the cassava mill effluent (CME) could be attributed to the contents of the effluents. CME is known to be high in organic carbon, which could then reduce the C/N ratio of impacted soil but increased the TOC. Shanhinroknsar et al, (2008) and Nattipong and Alissara (2006) agreed that soil impaction with organic matter results in decreased C/N ratio especially if the impacting material has low N content. This agrees well with this study, CME has very

low protein but high carbohydrate contents hence the observed low C/N ratio.

The high cyanogenic potentials observed near the waste nit agreed with studies

observed near the waste pit agreed with studies of Aderiye et al. (2005) and Nwabueze and Odunsi (2007). These researchers had reported values of 2.91-4.11 and 2.17-5.0 mg HCN kg from cassava peels and tuber pastes respectively. The high cynogenic potential had been attributed to the high cynogenic glucosides (Linamarin and lotaustain) contained in cassava. The values obtained by Aderive et al. (2005) and Nwabueze and Odunsi (2007) do not differ much from the results obtained in this study, especially as it was cassava effluents that was studied too. The high change in pH and temperature very close to the waste pit could be attributed to the high reductive biochemical oxidative and transformations taking place there. The break down of organic matter in the effluent was exothermic, which caused the increase in temperature while the metabolism of the little protein content released ammonia (NH3. The ammonia dissolved in the available moisture to cause the reported increase in pH value. A similar observation had been reported by Nwaugo et al, (2008a) and Chinvere (2001) in similar studies on cassava effluent.

Observations in this study concerning metals suggest that cassava effluent did not cause much variation on heavy metals contents of the soil. Most of these metals (heavy and trace) had no change or very slight negligible change in concentrations. However, only the calcium content of the soil was significantly affected. This calcium change in soil impacted with cassava waste had been reported by Aderiye and Laleye (2003) and Adeyemo (2005). The observed increase could have resulted from the impact of the cassava mill effluent (CME)

. In biological indices analyzed, results showed that the bioloads of all the bacteria groups increased with distance away from the waste pit suggesting adverse growth conditions towards the pit. However, values obtained at 100m away were not statistically higher than those from control. This observation suggests that the high content of cassava mill effluent suppressed bacterial growth but at very low could be support bacterial concentrations growth. The high content of cyanogenic glycosides (linamarin and lotaustiallin) in the CME was not metabolized by many microorganisms but when in low quantity, could be easily metabolized by the few organisms capable of doing so, while other bacterial species depend on the non-toxic intermediates produced. Nwaugo et al (2008a) and Karin (2006) reported that organic matter when added to the soil in small concentrations, encouraged bacterial growth. This observation agreed well with this study. Conditions and nutrients at 100m sampling point could have been optimum for bacteria growth, hence the results obtained.

Several authors (Nwaugo et al, 2007, Pelczar et al, 2003, Karin 2006, Onycagba et al, 2008) have stated that total heterotrophic bacterial counts are in all cases higher than the other specific bacterial groups in the soil. Similarly, Nwaugo et al. (2008a) and Prescott et al, (2001) reported that very slight change in environmental factors affect nitrifying bacteria adversely. These observations are evident in this study. While THB were the most prevalent in all the soil samples analyzed, NB were trhe least. NB were the most adversely affected bacterial group in the study. Some bacterial

species in the specialized group (LB, NB and PSB) may equally be found in the THB, making the THB more abundant than any other group.

Results from the enzyme activities analyzed showed some of the biogeochemical transformations taking place in the soil more vividly. The results correlated well with the findings in the bioloads of various bacteria groups. Dehydrogenase, which is found in intact bacterial cell, had its highest activities in the 100m soil sample and lowest in the most impacted soil (pit edge). This was exactly the case of the THB bioload with 3.7x10<sup>4</sup> at the pit edge but 7.4 x 10° at the 100m sample spot. Olivera and Pampulha (2006), Lee et al, (2002) and Nwaugo et al (2008b) had reported a similar observation dehydrogenase. with dehydrogenase is found in living bacterial cells, it therefore follows that the more the bacterial cells, the higher the production and consequent activities of the enzyme.

The activities of urease were also significant in this study. Urease has its highest activities in the most impacted soil (pit edge) which had the highest TOC content and low N content. Shahinrokhsar et al, (2008) stated that high TOC stimulated urease activities while Cookson and Lepiece (1996) reported increased urease activities in N-content soil. The increased metabolism of TOC resulted in the release of high amount of NH<sub>3</sub> which impacted the characteristic urine (NH<sub>3</sub>) odour around cassava mills.

Activities of acid and alkaline phosphatase were in opposite direction. While alkaline phosphatase activities increased towards the pit edge that of acid phosphatase decreased in that direction but increased in the opposite. This observation agreed very well with the prevailing environmental conditions especially the pH. Soil pH towards the waste pit was alkaline. The break down of the little protein content in the effluent which resulted in the production of NH<sub>3</sub> and consequent increase in soil pH made conditions favourable for alkaline phosphatase activities but unfavourable for acid phosphatase. Alkaline phosphatase operated best at the pH of 11.5 while acid phosphatase acts better at pH

6.8. the optimum pH of activities of these enzymes could have been the major cause in the observed variations in the rates of activities especially as they had similar rates in the control-soil and 100m away soil.

In conclusion, this study revealed adverse environmental effects of cassava mileffluents on soil physicochemical and biologic parameters. Again, it also calls for serious rehabilitation, if the soil will be used for agricultural and other purposes as the factor important in soil health are negatively affected in addition, this study revealed absence of proper regulation in the disposal of wastes an industrial effluents and so calls for introduction of such regulations to control the disposal of wastes generated from such industries. The becomes more glaring as most of these cassaumills are sited near residential areas.

#### References

Aderiye, B.I and Laleye, S.A. (2003). Relevant of fermented food products in southwe Nigeria. *Plant foods Hum. Nutria* 58:1-1

Adewoye, S.O; Fawolc O.O.; Owplabi, O.D. Omotosho, J.S (2005) Toxicity of Cassa Waste Water Effluents to Africa Catfish, Clarias gariepinus-Burche 1822. Ethiop. J.Sc 28 (2): 180194.

Akani, N.P., Nmelo, S.A. and ihemanandu, l. (2006). Effects of cassava processi effluents on the microbial population aphysiochemical properties of loamy soll Nigeria. 10<sup>th</sup> Annal. Conf. Nig Soc. I microbial, Keffi, 10-14<sup>th</sup> October, 2006.

APHA, (19980. Standard methods examination of water and waste water edition. American Public Hot Association, Washington, D.C.

Cassida, L.E; Klein, J.O and Snatoro, D. (196) Soil Dehydrogenase activity Soil St 98:371-374.

Chinyere, G.C (2001). Effects of Effluents in cassava processing plants on soil cyallevels. A case study of Okigwe in State, Nigeria. J Hith Vis Sci. 3:38-93.

Chessbrough, m. (2003). Laboratory Manuelj Tropical countries Vol. 11. microbiology Tropical Ha Technology. ELBS London.

- K. (1985). A New potential for neglected crops. Westview Press, Boader Co. USA.
- enzyme activity of soils of the Bantinah rgon of Sultanate of Oman. J Arid. Environ. 32225-238.
- Desse, G and Tay M. (2001). Microbial loads and Microflora of cassava (Manihot esculenta Grantz) and effects of cassava juice on some food borne pathogens. J Food Tech. Afri. 6(1):21-24.
- Karin, N. (2002). Impact of organic waste residue on structure and function of soil bacterial communities. Ph.D Thesis. Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Kandeler, E. and Gerber, H. (1988). Short term assay of urease activity using colorimetric determination of ammononium salt. *Boil*. fert. Soils 6:68-72.
- Lee. I. Kin. K Chang, Y. Bac, B. Kin, H.H and Baek K.(2002) Heavy metal concentrations and enzyme activities in soil from a contaminated Korean shooting range. J. Biose. Biomin 94:406-411.
- Li. H. Zhang, y. Zhang, C.g. AND chen, G. X. (2005). Effects of petroleum containing waste water-irrigation on bacterial diversities and enzymatic activities in a paddy soil irrigation area. J. environ. Qual. 34:1073-1080.
- Nattipong, K and Alissara, K. (2006). Effects of cassava Pulp and swine manure compost on growing plants in greenhouse *J. Water Environ. Technol.* 4 (1):9-32.
- Nwabueze, T.U and Odunsi, F.o (2007).

  Optimization ofprocess conditions from cassava (manihot esculenta) Lafun production. Afri J. Biotech. 6(5):603-611.
- Nwaugo, V.O; Onyeagba, R.A.; Chima, G.N. and Umeham, S.N. (2007). Effects of discharge of petroleum produced water on the physicochemical and potential flora of Nkissa River.
- Egbema, Rivers State. Int. J. Biotechnol. Allied Sci. 2 (2):126-130.
- Nwaugo, V.O.; Chima, G.N. Ekot C.A and Umeham, S.N (2008a), effects of cassava effluents on the physiochemical parameters and microbial flora of Nkissa

- River, Egbema Rivers State. Int. J Biotechnol and Allied Sci. 3(1):346-353.
- Nwaugo, V.O; Chinyere, G.C. and Inyang, C.U. 2008). Effects of palm oil mill effluent (POME) on soil bacterial flora and enzyme activities in Egbema River State. *Plant Prod Res. J.* 12:10-3.
- Nwaugo, V.O; Onyeagba, R.A.; Akubugwo, I.E and Ugbogu, O.C (2008c). soil bacterial flora and enzymatic activites in heavy metals (Pb and Zn) contaminated soil for Ishiagu, Ebonyi state. *Biokemistri* 20(2):77-84.
- Oliveira, A and Pampulha, ME.E. (2006). Effects of Long-term heavy metal contamination on soil microbial characteristics. J. Biosc. Bioeng 102(b)157-101.
- Onyeagba, R.A; Nwaugo, C.E and Verla, A.W. (2008). Phytoremediation OF Pb and Zn Polluted soil with *Chromolana* odaranta and Talinum Triangulare in Ishiagu, Mining area, Ebonyistate University, Uturu, 10-13<sup>th</sup> October 2008.
- Oyewole, O.B and Odunfa, S.A. (1992). Characterization and distribution of lactic bacteria in cassava fermentation during fufu production. J. Appl. Bacterial, 68:148-152.
- Oyewole, O.B Afolami, O.A (2001). Quality and Preference of different cassava varieties for Lafun production. *J Food Tech. Afr.* 6:27-29.
- Prescott, L.M: Klein, D.A and harley, J.P. (2005). Microbiology. 6<sup>1H</sup> Edition. McGraw Hill Education Books. USA 316-386.
- Shanhinrokhsar, P.; Vahed, H.S. and Haghuadl A. (2008). Evaluation of paddy soils properties on urease enzymes activity. Conference on international research on Food security, National resource Management and Rural Development, University of Hoheheim, October, 7-9, 2008
- Tabatabai, M.A (1997). Soil enzyme. In: page.
  A.L Millers. R.H and Kenny D.R. (eds)
  Methods of soil analysis Part II: Chemical
  and Microbiological Properties. Soil
  Science society of America Madison W.I.

Taye, M. (1994). Cassava in Southern and Southwestern Ethiopia. Cassava News Letter, 18:6-9 CIAT. Columbia.

U.S. Patient Storm 6638730 (2003)
Composition for qualitative screening of phosphate solubilizing microorganisms Issued October, (microorganisms and qualitative 2003.